Detection of Group A Rotavirus in Faeces of Diarrhoeic Bovine Porcine and Human Population from Eastern India by Reverse Transcriptase–Polymerase Chain Reaction

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ABSTRACT

The present study describes the distribution of group A rotavirus in animals and human population of eastern state of India–West Bengal. During the study, a total of 211 samples were collected from diarrhoeic bovine, porcine and human population (below 6 months of age). Of which, 26 (12.32%) samples were found positive for group A rotavirus by VP6 gene based reverse transcriptase–polymerase chain reaction (RT–PCR) assay. A total of 89 bovine, 82 porcine faecal and 40 human stool samples were screened by RT–PCR assay for the detection of group A rotavirus, of which 10 bovine (11.23%), 9 porcine (10.97%) and 7 human (17.5%) samples were detected positive. The study revealed baseline information to understand rotavirus epizootology and future prophylactic strategies against rotavirus in eastern India.

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Rotaviruses are recognized as the major viral etiological agents causing diarrhoea in neonates of all species including animals, human and birds. Rotavirus accounts for more than 6 million deaths in infants worldwide (Estes and Kapikian, 2007), which are more in developing countries including India (Parasar et al., 2006). Rotaviruses have accounted for 25% mortality in young animals and causes severe economic losses in livestock sector (Fagiolo et al., 2005). The virus belongs to the genus Rotavirus under the family Reoviridae and group A rotaviruses are major pathogens causing acute gastroenteritis in children and animals (Ciarlet et al., 2002).

Combined studies on the prevalence of rotavirus associated diarrhoea in bovine and porcine species of animals and human have not been documented in eastern India. This study was aimed to understand the burden and epidemiology of rotavirus infection in West Bengal, one of the eastern states of India. The results of this study are expected to be considered as baseline information for future studies to determine genotypes and subsequent vaccine against rotavirus.

A total of 211 samples were collected from bovine (n=89), porcine (n=82) and human (n=40) (below the age of 6 months) affected with acute diarrhoea from different districts of West Bengal viz. Purulia (n=35), Burdwan (n=34), Nadia (n=36) and Kolkata (n=86). The samples were transported to the laboratory for processing using ice cold container (Thermoflask, Milton, India). The samples were stored at −20°C for extraction of viral RNA. A 10% (w/v) faecal suspension of the faecal material was prepared in phosphate buffer saline (PBS; pH 7.2). Centrifugation was carried out, after thorough vortexing, at 7500 rpm for 20 min to remove the coarse debris. Total RNA was extracted using RiboZol RNA extraction reagent (Amresco, USA) as per manufacturer instruction. The dsRNA was subjected to reverse transcription as per the method described by Iturriza–Gomara et al. (2004). The synthesized cDNA was stored at −20°C till further use. Detection of group A rotavirus, amplification of partial length VP6 gene was carried out using primers and conditions as optimized by Kattoor et al. (2013). The primer sequences and nucleotide position of oligonucleotide primers are shown in Table 1.

All the 211 samples including 89 bovine, 82 porcine faecal and 40 human stool samples were screened by the VP6 gene based RT–PCR assay. PCR was carried out by using primers for the group specific VP6 gene which generated expected amplicon size 227 bp (Figure 1). In total 12.32% (26/211) diarrhoeic faecal samples were positive for group A rotavirus. On RT–PCR analysis, 10 bovine (11.23%), 9 porcine (10.97%) and 7 human (17.5%) samples were found positive for the presence of group A rotavirus. The RT–PCR offers many advantages besides high sensitivity and specificity in detection of rotavirus in faecal samples (Fedorova et al., 2005; Kang et al., 2004). It helps in the detection of viral nucleic acid

Table 1. Primers used for partial length amplification of VP6 gene of Group A rotavirus

<table>
<thead>
<tr>
<th>Gene</th>
<th>Primers</th>
<th>Sequence (5´–3´)</th>
<th>Amplicon size (bp)</th>
</tr>
</thead>
<tbody>
<tr>
<td>VP6</td>
<td>GARV–D–</td>
<td>TTTGATCACTAAYT</td>
<td>227 bp</td>
</tr>
<tr>
<td>VP6F</td>
<td></td>
<td>ATTICACC</td>
<td></td>
</tr>
<tr>
<td>VP6</td>
<td>GARV–D–</td>
<td>GGTCACATCCCTCTC</td>
<td></td>
</tr>
<tr>
<td>R</td>
<td></td>
<td>ACTA</td>
<td></td>
</tr>
</tbody>
</table>

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during initial stages of infection without waiting for higher virus titre and development of immune response in the affected host species (Niture et al., 2010). Detection of rotavirus infection in reservoir animals and carriers is another advantage of RT–PCR (Niture et al., 2010). The PCR techniques are used throughout the world for the group A rotavirus typing in strains obtained directly from faecal extracts and/or cell culture (Basera et al., 2010; Mondal et al., 2011).

In India, Gulati et al. (1996) was the first to determine the genetic diversity of bovine group A rotavirus using RT–PCR. In a study by Basera and co-workers (2010), group A rotavirus was found in 7.81% (10/128) samples from bovine calves. Chinsangaram et al. (1995) detected group A rotavirus from 94% (44/47) calves, directly by RT–PCR in faecal samples. In the present study, we detected high incidence of rotavirus among human (17.5%), followed by bovine (11.23%) and porcine (10.97%) samples. The findings are in agreement with the report from Western India that revealed 20.25% of human rotaviruses followed by 12.9% of bovine and 7.84% of avian rotaviruses (Niture et al., 2011). Earlier, 9.73% incidence of bovine group A rotavirus was reported from Mumbai region (Western part of India) (Mondal et al., 2012). The differences in the prevalence of rotavirus could be some factors like hygienic measures, proper access to maternal colostrum, nutrition, season and climatic factors such as rainfall, temperature, relative humidity etc.

In conclusion, the group A rotavirus is prevalent in bovine and porcine species of animals and human in eastern Indian state, West Bengal. RT–PCR assay was evaluated as sensitive and specific assay for the rapid detection of group A rotavirus in faecal samples. In Indian subcontinent, human live in close proximity to their livestock, often in poor sanitary conditions, contamination of water and food is possible. It may be responsible for possible zoonotic transmission of rotaviruses. So, in order to obtain a much better perspective in terms of the circulating genotypes in different species and regions of the country, molecular epidemiological surveillance of the circulation of group A rotaviruses in varied host species needs to be carried out.

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REFERENCES


